High Rates of New Delhi Metallo-β-Lactamase Carbapenemase Genes in Multi-Drug Resistant Gram-Negative Bacteria in Nicaragua

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Abstract. Antimicrobial resistance (AMR) is a global public health crisis. Much of the burden of AMR in resourcelimited settings remains unknown. This pilot study characterized clinical isolates of multidrug-resistant Gram-negative rods (MDR-GNRs) from Nicaragua. New Delhi metallo-β-lactamase (NDM) carbapenemase genes were detected in 60% of isolates. Enterobacteriaceae had the highest rates of NDM detection, with 92% (50/54 isolates) positive by polymerase chain reaction (PCR). Pulsed-field gel electrophoresis (PFGE) analysis revealed patterns of clustering among isolates by two factors: plasmid profiles and year of culture. These findings of very high rates of NDM-carbapenemase genes in MDR-GNRs from hospitals throughout Nicaragua are alarming. Further research is needed to determine clinical and epidemiologic factors associated with multidrug-resistant isolates and to guide interventions to limit further spread.

Antimicrobial resistance (AMR) is a global public health crisis that requires coordinated efforts between countries and international organizations. Public health institutions, including the CDC and WHO, recognize AMR as one of the most important threats to public health of our time. Rates of multidrug-resistant (MDR) bacterial isolates, including extended-spectrum βlactamases (ESBLs), carbapenem-resistant Enterobacteriaceae, Acinetobacter baumannii, and Pseudomonas aeruginosa, are on the rise worldwide.^{1-[4](#page-3-0)}

The most common mechanism for development of multidrugresistance in Gram-negative bacteria is via horizontal transfer of plasmids carrying resistance genes, particularly carbapenemases. [5](#page-3-0) Carbapenemases are members of the molecular Class A, B, and D β-lactamases and can be organized further into enzyme families. 6 Class A and D β -lactamases work by a serinebased hydrolytic mechanism. Klebsiella pneumoniae carbapenemases (KPCs) are the most prevalent of the class A group, whereas oxacillin-hydrolyzing–type (OXA) carbapenemases are the most prevalent class D carbapenemases.^{[7](#page-3-0)} Class B enzymes are metallo-β-lactamases and include the VIM (verona integron-mediated metallo-β-lactamase), imipenem metallo-β-lactamase (IMP) (active on IMP), and New Delhi metallo-β-lactamase (NDM) families (among others). Carbapenemase gene detection is a critical component of surveillance at all levels—local, national, and international.

Much of the burden of AMR in resource limited settings remains unknown. In Nicaragua, no prior published studies have characterized AMR among clinical isolates. From 2014 to 2017, 334 Gram-negative rod (GNR) isolates were sent to Centro Nacional de Diagnóstico y Referencia (CNDR) for confirmation of carbapenemases; 268 were processed and 66 were nonviable (Supplemental Table 1). Isolate distribution by year is shown in Supplemental Table 1. Of the 268 isolates tested, 231 isolates were defined as being resistant to imipenem by disk diffusion. This pilot study examined 100

selected MDR-GNR isolates from throughout the country to determine the genetic relatedness of resistant strains and better characterize the mechanisms of resistance.

Isolates from sterile sites collected from 2014 to 2017 from hospitals throughout Nicaragua were originally sent to the CNDR in Managua. Isolates were obtained from patients ranging in age from 1 day old to 90 years old and from a variety of hospital departments, including the neonatal intensive care unit (ICU), pediatric and neonatal floors, and adult ICU. The majority of the isolates were cultured from blood (81%) with the remainder cultured from pleural fluid (7%), cerebrospinal fluid (CSF) (4%), peritoneal fluid (2%), bronchial lavage (2%), and source unknown (4%) (Supplemental Table 2). Isolates were identified at the CNDR using the Vitek 2 (bioMérieux, Marcy l'Etoile, France). Isolates with carbapenem resistance identified by E-test, with or without confirmed resistance to additional antibiotic classes, were stored in media supplemented with 25% glycerol. Hundred suspected multidrug-resistant Gram-negative rods (MDR-GNRs) representing the most common species (Escherichia coli, K. pneumoniae, A. baumannii, and P. aeruginosa) were shipped to Emory University for further characterization. Of the 100 isolates, 16% were P. aeruginosa, 30% were A. baumannii, and 54% were Enterobacteriaceae. The majority of Enterobacteriaceae were K. pneumoniae (50/54, 93%) and remainder were E. coli (4/54, 7%).

Susceptibility testing of all isolates to appropriate carbapenems for each species was performed by disk diffusion according to Clinical and Laboratory Standards Institute.^{[8](#page-3-0)} Resistance to carbapenems was confirmed for 99 of the 100 isolates. One P. aeruginosa isolate was found to be susceptible to all carbapenems tested. All isolates were found to be susceptible to colistin by E-test (Supplemental Table 2). Although colistin testing by gradient diffusion has limitations,^{[8](#page-3-0)} many clinical microbiology laboratories use this methodology. In addition, our isolates had very low minimum inhibitory concentrations (none bordering a resistant phenotype or colonies inside the zone of inhibition) and it is, therefore, unlikely that any resistant isolates were missed by this method (Supplemental Table 2).

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IABLE 1 Summary of findings for all isolates					
Ertapenem	Doripenem	Imipenem	Producer ⁺	Producer (%)‡	
Klebsiella pneumonia, $n = 50$	49 (98%) \$	48 (96%) \$	49 (98%) \$	21 (42%)	50 (100%)
Escherichia coli, $n = 4$	4 (100%)	4 (100%)	4 (100%)	$0(0\%)$	4 (100%)
Pseudomonas aeruginosa, $n = 16$	ND	15(94%)	14 (87%)	ND	11 (68.8%)
Acinetobacter baumannii, $n = 30$	ND	30 (100%)	30 (100%)	ND	l (3%)¶

TABLE 1

ND = not tested. * Determined by disk diffusion.

† Determined using a combination disk diffusion method (ceftazidime + ceftazidime–clavulanate and cefotaxime + cefotaxime–clavulanate).

‡ Determined by the modified carbapenem inactivation method (mCIM) phenoptypic test for the production of carbapenemase.

§ Remaining isolates were intermediate to ertapenem, doripenem, and meropenem.

One P. aeruginosa was susceptible to all carbapenems tested and one was intermediate to imipenem.

{ Note: mCIM has been found to be less accurate for A. baumannii.

Enterobacteriaceae isolates were tested for the presence of ESBLs using a combination disk diffusion method (ceftazidime + ceftazidime-clavulanate and cefotaxime + cefotaxime-clavulanate).[9](#page-3-0) Presence of an ESBL was found in 21/50 (42%) of the K. pneumoniae isolates, but none of the E. coli isolates. All isolates (Enterobacteriaceae, A. baumannii, and P. aeruginosa) were tested for carbapenemase production by the modified carbapenem inactivation method (mCIM).^{[9](#page-3-0)} All 54 Enterobacteriaceae, 11/16 (68%) P. aeruginosa, and 1/30 (3%) of A. baumannii were found to produce a carbapenemase by the mCIM. Results are shown in Table 1 and Supplemental Table 2.

Bacterial DNA was extracted using an eMag® instrument (bioMérieux), and all isolates were screened for the presence of β-lactamase resistance genes bla_{KPC}, bla_{OXA-48}, bla_{VIM}, bla_{IMP}, and bla_{NDM} as previously described.^{10,11} Table 2 details the distribution of carbapenemase genes by each bacterial species. All Enterobacteriaceae isolates carried a carbapenemase gene with the vast majority 50/54 (93%) being positive for the bla_{NDM}; 46/50 (92%) K. pneumoniae and 4/4 (100%) E. coli. The bla_{KPC} was found in 4/50 (8%) of the K. pneumoniae isolates. Acinetobacter baumannii and P. aeruginosa isolates were less likely to have a carbapenemase gene identified compared with the Enterobacteriaceae; 11/16 (68.8%) of the P. aeruginosa and 10/30 (33%) of A. baumannii isolates carried a carbapenemase gene. Pseudomonas aeruginosa isolates carried bla_{VIM} and b/a_{IMP} carbapenemase genes only, with the majority of these isolates (10/11, 91%) having both. The 10/30 (33%) of A. baumannii isolates with a carbapenemase gene were identified as bla_{NDM} PCR confirmed the results of the mCIM for all Enterobacteriaceae and P. aeruginosa isolates. The mCIM results were less reliable for use with A. baumannii isolates as seen by others.¹²

Genetic relatedness and plasmid sizes were determined by PFGE using the CHEF-Mapper (Bio-Rad, Hercules, CA) and analyzed with BioNumerics v. 7.6 (Applied Maths, Saint-

Martens-Latem, Belgium).^{[13](#page-3-0)–[16](#page-3-0)} PFGE analysis revealed patterns of clustering among isolates. Isolates appeared to cluster by three factors: plasmid profiles, year of culture, and health care setting ([Figure 1](#page-2-0)). Clustering was particularly apparent for K. pneumoniae isolates ([Figure 1A](#page-2-0)); however, more plasmids were identified within these strains.

This study reveals very high rates of NDM-carbapenemase genes in MDR-GNRs from hospitals throughout Nicaragua. Prior studies have shown the presence of NDM-1 in Latin and Central America, but there is limited evidence regarding the prevalence of this resistance mechanism in most of Central America, and particularly in Nicaragua.^{[17](#page-3-0)-[19](#page-3-0)} The burden of MDR-GNR may be underestimated as approximately 38% of the isolates were either nonviable or not received by the CNDR. PFGE analysis revealed clusters of isolates by year of isolation, health care setting, and approximate plasmid size, suggesting genetic relatedness. Available clinical data for the 100 tested isolates were limited. However, only strains isolated from sterile sites were selected for this study, indicating the clinical relevance of these data.

Further research is needed to examine clinical and epidemiologic factors associated with MDR isolates and to determine patterns of acquisition, as well as high-risk exposures or comorbidities associated with MDR infection in Nicaragua. In settings with limited health care resources, cost-effective management and surveillance strategies are warranted to control transmission and provide appropriate empiric therapy to individuals with bacterial infections to improve clinical outcomes. This study allows us to draw conclusions about the prevalence of certain types of carbapenemase among MDR-GNR isolates; however, further research is needed to determine the overall prevalence of MDR-GNR. A prospective study that collects epidemiologic and geographic information for isolates could help identify specific regions and populations within Nicaragua with higher rates of community MDR carriage. Determining the burden of highly resistant bacteria in Nicaragua is a crucial first step to guide interventions to limit further spread both locally and globally.

Received July 28, 2019. Accepted for publication October 14, 2019.

Published online November 25, 2019.

Note: Supplemental tables appear at www.ajtmh.org.

Acknowledgment: We thank Janet Thonkulpitak for her help with the PFGE.

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FIGURE 1. Cluster analysis of PFGE patterns (data not shown), carbapenemases, and plasmids of Enterobacteriaceae (A), Pseudomonas aeruginosa (B), and Acinetobacter baumannii (C) isolates from 2014 to 2017 (Yr) shown in red, blue, purple, and green, respectively. Percent similarity of PFGE bands at top of dendrogram. Size of plasmids is approximate kb. This figure appears in color at www.ajtmh.org.

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